

Durham Research Online

Deposited in DRO:

06 September 2018

Version of attached file:

Accepted Version

Peer-review status of attached file:

Peer-reviewed

Citation for published item:

Sun, H.-J. and Smith, M.R. and Zeng, H. and Zhao, F.-C. and Li, G.-X. and Zhu, M.-Y. (2018) 'Hyoliths with pedicles illuminate the origin of the brachiopod body plan.', *Proceedings of the Royal Society B : biological sciences.*, 285 (1887). p. 20181780.

Further information on publisher's website:

<https://doi.org/10.1098/rspb.2018.1780>

Publisher's copyright statement:

Additional information:

Use policy

The full-text may be used and/or reproduced, and given to third parties in any format or medium, without prior permission or charge, for personal research or study, educational, or not-for-profit purposes provided that:

- a full bibliographic reference is made to the original source
- a [link](#) is made to the metadata record in DRO
- the full-text is not changed in any way

The full-text must not be sold in any format or medium without the formal permission of the copyright holders.

Please consult the [full DRO policy](#) for further details.

1 **Hyoliths with pedicles illuminate the origin of the brachiopod body plan**

2 Haijing Sun^a, Martin R. Smith^{b,1}, Han Zeng^{a,c,d}, Fangchen Zhao^{a,1}, Guoxiang Li^a,
3 Maoyan Zhu^{a,c}

4 ^aState Key Laboratory of Palaeobiology and Stratigraphy, Nanjing Institute of
5 Geology and Palaeontology and Center for Excellence in Life and Palaeoenvironment,
6 Chinese Academy of Sciences, No. 39 East Beijing Road, Nanjing 210008, China

7 ^bDepartment of Earth Sciences, Mountjoy Site, Durham University, South Road,
8 Durham DH1 3LE, UK

9 ^cCollege of Earth Sciences, University of Chinese Academy of Sciences, No. 19
10 Yuquan Road, Beijing 100049, China

11 ^dDepartment of Paleobiology, National Museum of Natural History, P.O. Box 37012,
12 MRC-121, Washington, DC, 20013–7012, USA

13 ¹To whom correspondence should be addressed. Email: fczhao@nigpas.ac.cn and
14 martin.smith@durham.ac.uk

15 **Abstract**

16 Hyoliths are a taxonomically problematic group of Palaeozoic lophotrochozoans that
17 are among the first shelly fossils to appear in the Cambrian period. On the basis of
18 their distinctive exoskeleton, hyoliths have historically been classified as a separate
19 phylum with possible affinities to the molluscs, sipunculans or lophophorates – but
20 their precise phylogenetic position remains uncertain. Here we describe a new
21 orthothecide hyolith from the Chengjiang Lagerstätte (Cambrian Series 2 Stage 3),
22 *Pedunculotheca diania* Sun, Zhao et Zhu gen. et sp. nov., which exhibits a
23 non-mineralized attachment structure that strikingly resembles the brachiopod pedicle
24 – the first report of a peduncular organ in hyoliths. This organ establishes a sessile,
25 suspension feeding ecology for these orthothecides, and – together with other
26 characteristics (e.g., bilaterally symmetrical bivalve shell enclosing a filtration
27 chamber and the differentiation of cardinal areas) – identifies hyoliths as stem-group
28 brachiopods. Our phylogenetic analysis indicates that both hyoliths and crown
29 brachiopods derived from a tommotiid grade, and that the pedicle has a single origin
30 within the brachiopod total group.

31 **Keywords:** Pedicle, orthothecide hyoliths, phylogeny, stem brachiopod, Chengjiang
32 Lagerstätte, Cambrian

33 1. Introduction

34 Hyoliths – orthothecides and hyolithides – are enigmatic Palaeozoic fossils known
35 principally from their originally aragonitic [1] shelly elements, which comprise an
36 elongate conical shell (‘conch’) capped with a lid-like operculum and, in the
37 hyolithide subgroup, a pair of elongate ‘helens’ [2]. On account of this unfamiliar
38 morphology, the ecology and relationships of this group have long attracted debate. A
39 recurrent suggestion treats hyoliths as an extinct phylum in taxonomic limbo between
40 molluscs and sipunculans [3], but recent reports of soft tissue anatomy [4] have led to
41 the disputed [5, 6] suggestion that hyoliths belong to the brachiozoan group, which
42 contains the brachiopod and phoronid phyla. However, their precise phylogenetic
43 position remains obscure.

44 We report a new, exceptionally preserved orthothecide genus from the early Cambrian
45 of China, which reveals taxonomically significant new details of the hyolith body plan.
46 Phylogenetic analysis of a new morphological dataset that encompasses the major
47 lophotrochozoan groups indicates that hyoliths are stem-group brachiopods, and
48 addresses longstanding palaeontological problems concerning the origin of the
49 brachiopod lineage.

50 2. Results

51 Superphylum Brachiozoa Cavalier-Smith, 1998

52 Stem-group of Phylum Brachiopoda Duméril, 1806

53 Class Hyolitha Marek, 1963

54 Order Orthothecida Marek, 1966

55 *Remarks.* Our specimens are attributed to Orthothecida (*sensu* [7]) based on the
 56 absence of a ligula and the absence of lateral sinuses (depressions of the commissure
 57 that accommodate the helens of hyolithides). Our material exhibits a highly
 58 compressed, subcircular cross-section, a pair of lateral furrows separating venter from
 59 dorsum, and one or two ventral grooves; as such, it cannot be accommodated in any
 60 existing orthothecide family. As the internal characters of the operculum and other
 61 essential features are not well preserved, we provisionally leave the family level
 62 taxonomy of this genus in open nomenclature.

63 ***Pedunculotheca diania* Sun, Zhao et Zhu gen. et sp. nov.**

64 1996 *Ambrolinevitus* Sysoyev, 1985; Chen *et al.*, 1996, p. 139, fig. 173 [8]

65 2004 *Ambrolinevitus ventricosus* Qian, 1978; Chen 2004, p. 203, fig. 308 [9]

66 (Figures 1–3; Supplementary figures S1 and S2)

67 **Etymology.** *Pedunculotheca* refers to the peduncular attachment structure of the shell;
 68 *diania* reflects an abbreviation of Yunnan Province, from where the fossils were
 69 recovered.

70 **Holotype.** NIGPAS (Nanjing Institute of Geology and Palaeontology, Chinese
 71 Academy of Sciences, Nanjing) 166593 (figure 1*a*).

72 **Additional material.** Twenty-four further slabs (NIGPAS 166594–166617), bearing
 73 44 conical shells and 17 opercula. Ten of the conical shells preserve incomplete
 74 attachment structures.

75 **Occurrence.** Cambrian Series 2 Stage 3, the Maotianshan Member of the Yu'an-shan
 76 Formation (*Eoredlichia-Wutingaspis* Assemblage Zone), at sections near Yaoying
 77 Village in Wuding County, Shankou Village in Anning County, and Ma'an-shan
 78 Village in Chengjiang County, Yunnan Province, China (see Supplementary table S1).

79 **Preservation.** The material exhibits Burgess Shale-type preservation of
 80 non-mineralized tissues [10] in the fashion typical of the Chengjiang deposits [11].

81 **Terminology.** Our description uses standard terminology (see [2, 12]), though we
 82 prefer the taxonomically neutral term 'conical shell' to 'conch' (following [4]).

83 **Diagnosis.** Orthothecide with slender, orthoconic, transversely ridged conical shell.
 84 Dorsum broadly rounded with a pair of lateral furrows, grading into broadly rounded

lateral margins. Venter similarly convex with one medial or two lateral furrows. Dorsoventral transition broadly rounded, producing a subcircular transverse section. Apex of conical valve bears rimmed subcircular disc in smaller specimens, regularly striated attachment structure with spinose terminal holdfast in larger specimens. Operculum circular to subcircular, bearing concentric growth lines, with bilobate bulge on exterior surface. Indistinct boundary between small cardinal shield and large conical shield on internal surface.

Description. Complete specimens range from 6.2 to 25.5 mm long (mean: 14.7 mm; n = 15), 2.4 to 5.5 mm in apertural width (mean: 4.2 mm; n = 12), and 2.3 to 5.6 mm in opercular diameter (mean: 3.1 mm; n = 9). The ratio of apertural width to shell length ranges from 0.21 to 0.43 (mean: 0.31; n = 12). The operculum shows no ontogenetic variation: in all specimens it is circular (figure 2*d*) and covered by concentric lines (figure 1*a* and *c*). The opercular diameter is slightly smaller than the apertural width, indicating that the operculum could be retracted a short distance within the conical shell (figures 1*a*, 2*c* and *e*). The external surface of the operculum is convex close to the mediodorsal part, and features a bilaterally disposed pair of ridges that emerge from a raised quadrangular or oval area near the summit (figures 1*a*, *c* and 2*c–e*). Internally, a sharp change in convexity distinguishes the dorsal and lateral regions of the operculum (figure 2*d*), weakly delimiting a small cardinal shield from a large conical shield.

105 The apex of the conical shell bears a flat circular region around 0.7 mm across,
 106 presumably corresponding to a metamorphic shell (*sensu* [13]). In small individuals,
 107 the apices are flattened, with no obvious opening. The flattened surface is surrounded
 108 by a prominent rim, rather than the recessed groove that would be expected if it
 109 represented a septum [14]. In larger specimens, this region is replaced by a
 110 non-biomineralized attachment structure, measuring 2.4 mm in length where it is
 111 unambiguously complete (figure 1*a*). This stalk-like structure bears a central cavity
 112 (figures 1*c*, *d*, 2*a* and *b*) and an external ornament of transverse striations whose
 113 spacing and relief is consistent with the ridges that ornament the conical shell (figure
 114 1*a* and *b*): the spacing on attachment structure is 35 to 46 μm (mean = 38 μm ; $n = 30$);
 115 on the shell, 33 to 57 μm (mean = 44 μm ; $n = 240$). The structure terminates with a
 116 broad holdfast disk, from which marginal spines emerge (figure 1*a* and *b*). In adult
 117 specimens, the apical portion of the conical shell is triangular in dorsal profile, and
 118 exhibits a larger divergence angle (52–73°; mean: 59°; $n = 7$) than the rest of the shell
 119 (12–15°; mean: 13°; $n = 9$). Two smaller individuals without peduncular structures
 120 (figure 2*f* and *g*) suggest that these juveniles were attached by an apical part of the
 121 shell or an epithelium emerging from its apex. As the metamorphic shell is not present
 122 in larger specimens, subsequent growth of the attachment structure through ontogeny
 123 must have been accommodated by resorption or detachment of the apex, as in certain
 124 hyoliths (e.g. [14]) and brachiopods (e.g. [15]).

Ecology. The *Pedunculotheca* attachment structure (figure 3) seems to have been used for anchorage, as indicated by its digitate holdfast. This implies a sessile habit, which is difficult to reconcile with the deposit feeding strategy typically reconstructed on the basis of a meandering, often infilled alimentary tract [16-18]. Meandering guts, however, are known from suspension feeders [19], so do not provide a decisive ecological signal. We therefore consider *Pedunculotheca*, like certain hyolithides [4, 16, 20], to have been a suspension feeder.

3. Affinity of hyoliths

The discovery of a peduncular structure in hyoliths is invaluable in resolving the affinity of this problematic group. Although attachment apparatuses are common and diverse in sedentary animals (e.g., [21]), a pedicle-like structure contributes to a suite of skeletal and anatomical structures that together indicate an affinity with brachiozoans.

To test this proposal and evaluate the evolutionary implications of our observations, we constructed a phylogenetic dataset of 54 Lophotrochozoan taxa, each scored for 225 morphological characters (Supplementary Information §1). Figure 4 depicts a consensus of trees recovered by parsimony analysis under equal and implied weights ($3 \leq k \leq 24$), after correcting for the impact of inapplicable data [22, 23], which

143 account for 18.5% of the non-ambiguous tokens. Details of tree search procedures,
144 and results of analysis under a Bayesian implementation of the Mk model [24, 25] and
145 the uncorrected Fitch algorithm [26], are provided in Supplementary Information §2–
146 4; the choice of method impinges on certain details of the analytical results, but does
147 not affect our main conclusions.

148 Our data indicate that hyoliths are monophyletic; brachiopods and hyoliths are
149 derived from a paraphyletic tommotiid grade [27, 28] (figure 4). Hyoliths are the
150 sister taxon to a clade containing the tommotiids *Micrina*, *Mickwitzia* and
151 *Heliomedusa*, plus the brachiopod crown group.

152 A sister-group relationship between hyoliths and crown-group brachiopods is
153 supported by characters including a bivalved shell arrangement enclosing a filtration
154 chamber (character 72) and the differentiation of cardinal areas (pseudointerareas)
155 (characters 92, 107). Hyoliths are excluded from the brachiopod crown group as they
156 lack a subset of brachiopod synapomorphies: impressions of an attachment structure
157 and setae on the larval shell; a low ventral interarea or pseudointerarea (secondarily
158 increased in lingulellotretids, acrotretids and some craniiforms) and a lophophore that
159 coils anteriad (rather than posteriad) [4].

160 This position of hyoliths close to the basal node of Brachiopoda, in the context of
161 outgroup taxa, resolves the polarity of certain characters within the brachiopod

162 lineage, clarifying the position of some otherwise enigmatic brachiopod groups [29].
163 We recognize paterinids, *Salanygolina*, chileids and kutorginates as successive stem
164 groups to the rhynchonelliforms (figure 4). The weakly-mineralized lingulellotretid
165 *Lingulosacculus*, interpreted as a link between phoronids and linguliforms [30],
166 instead falls within a monophyletic Linguliformea. The agglutinated *Yuganotheca* [31]
167 seemingly represents a tommotiid, and belongs to the brachiopod stem group (though
168 this last result is contingent on the correct handling of inapplicable data, see
169 Supplementary Information §3).

170 Deeper in the tree, the tommotiids *Eccentrotheca* and *Dailyatia* group with *Halkieria*
171 as a grade within total-group brachiopods from which hyoliths and crown-group
172 brachiopods were derived, in line with many previous proposals (e.g., [32, 33]).
173 Bayesian and Fitch parsimony, perhaps influenced by their mishandling of
174 inapplicable data [23], prefer trees that group these taxa closer to the molluscs; but
175 under both interpretations, tommotiids are reconstructed as a paraphyletic grade from
176 which hyoliths and brachiopods evolved.

177 Despite the excellent fossil record of brachiopods [29], a disparate array of
178 hypotheses have been put forwards to explain the origin of the brachiopod body plan
179 [28, 31, 33, 34]. Our new observations and phylogenetic framework shed light on the
180 origins of fundamental morphological innovations in the brachiopod total group.

181 Firstly, it is most parsimonious to reconstruct a single origin of the brachiopod pedicle
 182 stem-wards of hyoliths and the attached tommotiid *Paterimitra* (Supplementary
 183 Information §5.5). This corroborates the homology between the *Pedunculotheca*
 184 attachment structure and the brachiopod pedicle, indicating a primitively attached
 185 rather than vagrant ancestry of the brachiozoan lineage (cf. [35]).

186 It has been argued that the pedicles of linguliforms and rhynchonelliforms are not
 187 homologous [36-38], in part because they are secreted by different parts of the
 188 organism (the ventral mantle in linguliforms [39]; a larval pedicle lobe in living
 189 rhynchonelliforms [40]). Our results instead imply a stepwise transformation from an
 190 ancestrally linguliform-like pedicle to the derived state of extant rhynchonelliforms:
 191 the pedicle migrated from the hinge to the umbo in early-diverging *Salanygolina* [15]
 192 and chileids [41] (character 120), but retained its coelomic cavity (character 32) until
 193 the kutorginates had diverged. On this basis, we interpret the two pedicle openings in
 194 *Nisusia* [36] as representing an ontogenetic relocation of the pedicle.

195 The migration of pedicle to the apex of the ventral valve occurred independently in
 196 the linguliforms, by the enrolling of the delthyrium [42] (character 113); and again in
 197 *Pedunculotheca*. It is more parsimonious to accommodate this variation as
 198 modification to an existing pedicle than to infer multiple separate origins of this
 199 organ.

200 Secondly, our results illuminate the high variability of shell mineralogy within the
 201 brachiopods (character 128), which are conventionally subdivided based on the
 202 composition of the shell, i.e. phosphatic Linguliformea and calcareous
 203 Rhynchonelliformea or Craniiformea [43]. Our results show that this emphasis on
 204 mineralogy over morphology has led to the mis-classification of the phosphatic
 205 paterinids – which are not linguliforms, but early-diverging rhynchonelliforms that
 206 inherited a phosphatic mineralogy from the brachiopod common ancestor – and the
 207 calcitic *Mummpikia*, which is not a rhynchonelliform but an independently calcifying
 208 linguliform, as its morphology suggests [44] (figure 4). To this list we add three
 209 further instances of mineralogical modification: hyoliths represent an independent
 210 innovation of an aragonitic mineralogy, as do the trimerellids; and craniiforms
 211 obtained their calcitic shells independently from the rhynchonelliforms (as proposed
 212 by [44]) , from a trimerellid precursor. In sum, brachiopods subvert the general
 213 expectation that lineages rarely modify their mineralogy [45, 46].

214 Taking this further, our results show that shell microstructure is highly variable within
 215 the brachiopod total group. In particular, the ‘canaliculate’ microstructure (character
 216 137) [1, 44, 47, 48] that has been afforded great importance – even to the point of
 217 identifying the Ediacaran reef-dweller *Namacalathus* [49] as a brachiozoan – turns
 218 out to have multiple origins across the brachiozoan total group, as do the broader
 219 ‘punctae’ (character 138). Hyoliths have been argued to have a mollusc-like

220 mineralogy and microstructure [6]. We contend that these variable characteristics are
 221 easily reconciled with a brachiopod affinity, and need not arise through common
 222 ancestry with molluscs – a group that have no obvious analogue for hyolith traits such
 223 as the pedicle, lophophore, or paired bilaterally symmetrical shells, and whose
 224 distinctive synapomorphies – a radula and creeping foot – have not been identified in
 225 even the best-preserved hyoliths.

226 Identifying hyoliths as stem-group brachiopods also resolves character polarity within
 227 the group itself. Moysiuk *et al.* [4] regarded hyolithides as primitive within Hyolitha,
 228 speculatively equating helens with tommotiid sclerites. We instead identify
 229 *Pedunculotheca* as primitive within Hyolitha (figure 4), on the basis of its retained
 230 brachiopod-like features. Hyolithides and other orthothecides are united by the
 231 secondary loss of a pedicle (character 26) and the morphology of their metamorphic
 232 shell (character 2) [2, 50-53], whose small fusiform to globular aspect differs from the
 233 discoidal shape of the larval shells of *Pedunculotheca* and Cambrian brachiopods
 234 [54].

235 The recognition of hyoliths as stem brachiopods indicates a profound change in the
 236 character of the group at the close of the Cambrian period. Early in the Cambrian, the
 237 brachiopod lineage was dominated by tommotiids, linguliforms and hyoliths: taxa
 238 with a high metabolic overhead, reflected by the extensive gut and relatively high
 239 volume of metabolically active tissue [4, 55-57], the use of phosphate as a biomineral

[58], and in certain tommotiids, an apparently vagrant ecology [32]. The end of the Cambrian period witnessed a marked decline in these groups (Supplementary figure S3), with a corresponding explosion in the taxonomic diversity of anatomically modern crown-group brachiopods – in particular the rhynchonelliforms, whose narrow disparity [55] reflects a fundamentally distinct ecological strategy, with a hyper-efficient physiology adapting the lineage to nutrient-limited settings [59]. From this perspective, the decline in hyolith diversity after their mid-Cambrian zenith mirrors a broader decline in high-metabolism stem-group brachiopods, and their supplanting in less nutrient-rich environments by their rhynchonelliform counterparts.

Ethics. No special permissions were required for the collection of samples.

Data accessibility. Supplementary figures and tables, including details of phylogenetic analysis and results, are provided at

<https://web.archive.org/web/ms609.github.io/hyoliths/>; source code is available at

<https://github.com/ms609/hyoliths>.

Phylogenetic data is accessioned with MorphoBank, project number 2800:

<http://morphobank.org/permalink/?P2800>

Authors' contributions. H.S., F.Z. and M.Z. designed the research and M.Z. and F.Z. provided material; H.S., M.R.S. and H.Z. performed the research; M.R.S., H.S. and

G.L. contributed to the phylogenetic dataset; M.R.S. performed the phylogenetic analysis with input of H.S.; H.S. wrote the original draft; M.R.S. and H.S. revised and finalized the manuscript; all co-authors jointed the research and contributed to discussions.

Competing interests. The authors declare no competing interests.

Funding. This research was supported by funding the Strategic Priority Research Program (B) of the Chinese Academy of Sciences (XDB18000000), the National Natural Science Foundation of China (41602002, 41472012), and NSFC-RCUK_NERC joint (41661134048).

Acknowledgements. We thank Prof. Junyuan Chen for his permission to access his collections. Prof. Dave Harper, Prof. Douglas Erwin, Dr. Cédric Aria and Dr. Zongjun Yin are acknowledged for their helpful discussions and suggestions.

References

- [1] Kouchinsky, A.V. 2000 Skeletal microstructures of hyoliths from the Early Cambrian of Siberia. *Alcheringa* **24**, 65-81. (doi:10.1080/03115510008619525).
- [2] Martí Mus, M., Jeppsson, L. & Malinky, J.M. 2014 A complete reconstruction of

- the hyolithid skeleton. *Journal of Paleontology* **88**, 160-170. (doi:10.1666/13-038).
- [3] Malinky, J.M. & Yochelson, E.L. 2007 On the systematic position of the Hyolitha (Kingdom Animalia). *Memoirs of the Association of Australasian Palaeontologists* **34**, 521-536.
- [4] Moysiuk, J., Smith, M.R. & Caron, J.-B. 2017 Hyoliths are Palaeozoic lophophorates. *Nature* **541**, 394-397. (doi:10.1038/nature20804).
- [5] Kimmig, J. & Pratt, B.R. 2018 Coprolites in the Ravens Throat River Lagerstätte of northwestern Canada: implications for the Middle Cambrian food web. *Palaios* **33**, 125-140. (doi:10.2110/palo.2017.038).
- [6] Moore, J.L., Porter, S.M. & Sigwart, J. 2018 Plywood-like shell microstructures in hyoliths from the middle Cambrian (Drumian) Gowers Formation, Georgina Basin, Australia. *Palaeontology* **61**, 441-467. (doi:10.1111/pala.12352).
- [7] Marek, L. 1966 New hyolithid genera from the Ordovician of Bohemia. *Časopis Národního Muzea* **135**, 89-92.
- [8] Chen, J.Y., Zhou, G.Q., Zhu, M.Y. & Yeh, K.Y. 1996 *The Chengjiang Biota: A Unique Window of the Cambrian Explosion*. Taichung, National Museum of Natural Science; 222 p.
- [9] Chen, J.Y. 2004 *The dawn of animal world*. Nanjing, Jiangsu Science and Technology Press; 366 p.
- [10] Butterfield, N.J. 1995 Secular distribution of Burgess-Shale-type preservation. *Lethaia* **28**, 1-13. (doi:10.1111/j.1502-3931.1995.tb01587.x).

- 297 [11] Hou, X.-G., Siveter, D.J., Siveter, D.J., Aldridge, R.J., Cong, P.-Y., Gabbott, S.E.,
 298 Ma, X.-Y., Purnell, M.A. & Williams, M. 2017 *The Cambrian fossils of Chengjiang,*
 299 *China: The flowering of early animal life*. Second edition ed, Wiley-Blackwell; 316 p.
- 300 [12] Martí Mus, M. & Bergström, J. 2007 Skeletal microstructure of helens, lateral
 301 spines of hyolithids. *Palaeontology* **50**, 1231-1243.
 302 (doi:10.1111/j.1475-4983.2007.00700.x).
- 303 [13] Popov, L.E., Bassett, M.G., Holmer, L.E., Skovsted, C.B. & Zuykov, M.A. 2010
 304 Earliest ontogeny of Early Palaeozoic Craniiformea: implications for brachiopod
 305 phylogeny. *Lethaia* **43**, 323-333. (doi:10.1111/j.1502-3931.2009.00197.x).
- 306 [14] Skovsted, C.B., Pan, B., Topper, T.P., Betts, M.J., Li, G. & Brock, G.A. 2016 The
 307 operculum and mode of life of the lower Cambrian hyolith *Cupithec*a from South
 308 Australia and North China. *Palaeogeography, Palaeoclimatology, Palaeoecology* **443**,
 309 123–130. (doi:10.1016/j.palaeo.2015.11.042).
- 310 [15] Holmer, L.E., Stolk, S.P., Skovsted, C.B., Balthasar, U. & Popov, L. 2009 The
 311 enigmatic Early Cambrian *Salanygolina* – a stem group of rhynchonelliform chileate
 312 brachiopods? *Palaeontology* **52**, 1-10. (doi:10.1111/j.1475-4983.2008.00831.x).
- 313 [16] Marek, L., Parsley, R.L. & Galle, A. 1997 Functional morphology of hyolithids
 314 based on flume studies. *Věstník Českého geologického ústavu* **72**, 351–358.
- 315 [17] Devaere, L., Clausen, S., Alvaro, J.J., Peel, J.S. & Vachard, D. 2014 Terrenewian
 316 orthothecid (Hyolitha) digestive tracts from northern Montagne Noire, France;
 317 taphonomic, ontogenetic and phylogenetic implications. *PLoS One* **9**, e88583.

- (doi:10.1371/journal.pone.0088583).
- [18] Runnegar, B., Pojeta, J., Morris, N.J., Taylor, J.D., Taylor, M.E. & McClung, G. 1975 Biology of the Hyolitha. *Lethaia* **8**, 181-191. (doi: 10.1111/j.1502-3931.1975.tb01311.x)
- [19] Morton, J.E. 1960 The functions of the gut in ciliary feeders. *Biological Reviews* **35**, 92-139. (doi: /10.1111/j.1469-185X.1960.tb01324.x)
- [20] Butterfield, N.J. 2003 Exceptional fossil preservation and the Cambrian Explosion. *Integrative and Comparative Biology* **43**, 166-177. (doi: 10.1093/icb/43.1.166)
- [21] Lei, Q.P., Han, J., Ou, Q. & Wan, X.Q. 2014 Sedentary habits of anthozoa-like animals in the Chengjiang Lagerstätte: Adaptive strategies for Phanerozoic-style soft substrates. *Gondwana Research* **25**, 966-974. (doi:10.1016/j.gr.2013.01.007).
- [22] Smith, M.R. 2018 TreeSearch: phylogenetic tree search using custom optimality criteria. *The Comprehensive R Archive Network*. (doi:10.5281/zenodo.1042590)
- [23] Brazeau, M.D., Guillerme, T. & Smith, M.R. Accepted. An algorithm for morphological phylogenetic analysis with inapplicable data. *Systematic Biology*; pre-print at *bioRxiv*. (doi:10.1101/209775).
- [24] Lewis, P.O. 2001 A likelihood approach to estimating phylogeny from discrete morphological character data. *Systematic biology* **50**, 913-925. (doi:10.1080/106351501753462876).
- [25] Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D.L., Darling, A., Höhna, S.,

- 339 Larget, B., Liu, L., Suchard, M.A. & Huelsenbeck, J.P. 2012 MrBayes 3.2: efficient
 340 Bayesian phylogenetic inference and model choice across a large model space.
 341 *Systematic Biology* **61**, 539-542. (doi:10.1093/sysbio/sys029).
- 342 [26] Fitch, W.M. 1971 Toward defining the course of evolution: minimum change for
 343 a specific tree topology. *Systematic Biology* **20**, 406–416. (doi:
 344 10.1093/sysbio/20.4.406)
- 345 [27] Holmer, L.E., Skovsted, C.B. & Williams, A. 2002 A stem group brachiopod
 346 from the Lower Cambrian: support for a *Micrina* (halkieriid) ancestry. *Palaeontology*
 347 **45**, 875-882. (doi: 10.1111/1475-4983.00265)
- 348 [28] Skovsted, C.B., Brock, G.A., Topper, T.P., Paterson, J.R. & Holmer, L.E. 2011
 349 Scleritome construction, biofacies, biostratigraphy and systematics of the tommotiid
 350 *Eccentrotheca helenia* sp. nov. from the Early Cambrian of South Australia.
 351 *Palaeontology* **54**, 253-286. (doi:10.1111/j.1475-4983.2010.01031.x).
- 352 [29] Harper, D.A.T., Popov, L.E., Holmer, L.E. & Smith, A. 2017 Brachiopods: origin
 353 and early history. *Palaeontology* **60**, 609-631. (doi:10.1111/pala.12307).
- 354 [30] Balthasar, U. & Butterfield, N.J. 2009 Early Cambrian “soft-shelled”
 355 brachiopods as possible stem-group phoronids. *Acta Palaeontologica Polonica* **54**,
 356 307-314.
- 357 [31] Zhang, Z.F., Li, G.X., Holmer, L.E., Brock, G.A., Balthasar, U., Skovsted, C.B.,
 358 Fu, D.J., Zhang, X.L., Wang, H.Z., Butler, A., et al. 2014 An early Cambrian
 359 agglutinated tubular lophophorate with brachiopod characters. *Scientific Reports* **4**,

- 360 4682. (doi:10.1038/srep04682).
- 361 [32] Skovsted, C.B., Betts, M.J., Topper, T.P. & Brock, G.A. 2015 The early
 362 Cambrian tommotiid genus *Dalmanella* from South Australia. *Memoirs of the*
 363 *Association of Australasian Palaeontologists* **48**, 1-117.
- 364 [33] Zhao, F., Smith, M.R., Yin, Z., Zeng, H., Li, G. & Zhu, M. 2017 *Orthozanclus*
 365 *elongata* n. sp. and the significance of sclerite-covered taxa for early trochozoan
 366 evolution. *Scientific Reports* **7**, 16232. (doi:10.1038/s41598-017-16304-6).
- 367 [34] Altenburger, A., Martinez, P., Budd, G.E. & Holmer, L.E. 2017 Gene expression
 368 patterns in brachiopod larvae refute the "brachiopod-fold" hypothesis. *Frontiers in*
 369 *Cell and Developmental Biology* **5**, 74. (doi:10.3389/fcell.2017.00074).
- 370 [35] Budd, G.E. & Jackson, I.S. 2016 Ecological innovations in the Cambrian and the
 371 origins of the crown group phyla. *Philosophical Transactions of the Royal Society B:*
 372 *Biological Sciences* **371**, 20150287. (doi:10.1098/rstb.2015.0287).
- 373 [36] Holmer, L.E., Popov, L.E., Ghobadi Pour, M., Claybourn, T., Zhang, Z., Brock,
 374 G.A. & Zhang, Z. 2018 Evolutionary significance of a middle Cambrian (Series 3)
 375 in situ occurrence of the pedunculate rhynchonelliform brachiopod *Nisusia sulcata*.
 376 *Lethaia* **51**, 424-432. (doi:10.1111/let.12254).
- 377 [37] Carlson, S.J. 1995 Phylogenetic relationships among extant brachiopods.
 378 *Cladistics* **11**, 131–197. (doi:10.1111/j.1096-0031.1995.tb00084.x).
- 379 [38] Carlson, S.J. 2016 The evolution of Brachiopoda. *Annual Review of Earth and*
 380 *Planetary Sciences* **44**, 409-438. (doi:10.1146/annurev-earth-060115-012348).

- 381 [39] Yatsu, N. 1902 On the development of *Lingula anatina*. *Journal of the College of*
 382 *Science, Imperial University, Tokyo, Japan* **17**, 1-112.
- 383 [40] Holmer, L.E., Zhang, Z., Topper, T.P., Popov, L. & Claybourn, T.M. 2017 The
 384 attachment strategies of Cambrian kutorginate brachiopods: the curious case of two
 385 pedicle openings and their phylogenetic significance. *Journal of Paleontology* **92**,
 386 33-39. (doi:10.1017/jpa.2017.76).
- 387 [41] Zhang, Z., Holmer, L.E., Popov, L. & Shu, D. 2011 An obolellate brachiopod
 388 with soft-part preservation from the Early Cambrian Chengjiang fauna of China.
 389 *Journal of Paleontology* **85**, 460-463. (doi:10.1666/10-121.1).
- 390 [42] Popov, L.E. 1992 The Cambrian radiation of brachiopods. In *Origin and early*
 391 *evolution of the Metazoa* (eds. J.H. Lipps & P.W. Signor), pp. 399-443. New York,
 392 Springer.
- 393 [43] Williams, A., Carlson, S.J., Brunton, C.H.C., Holmer, L.E., Popov, L.E., Mergl,
 394 M., Laurie, J.R., Bassett, M.G., Cocks, L.R.M., Rong, J.-Y., et al. 2000 *Brachiopoda*.
 395 *Linguliformea, Craniiformea, and Rhynchonelliformea*. in *Treatise on Invertebrate*
 396 *Paleontology*, The University of Kansas; 919 p.
- 397 [44] Balthasar, U. 2008 *Mummpikia* gen. nov. and the origin of calcitic-shelled
 398 brachiopods. *Palaeontology* **51**, 263–279. (doi:10.1111/j.1475-4983.2008.00754.x).
- 399 [45] Zhuravlev, A.Y. & Wood, R.A. 2008 Eve of biomineralization: Controls on
 400 skeletal mineralogy. *Geology* **36**, 923. (doi:10.1130/g25094a.1).
- 401 [46] Porter, S.M. 2010 Calcite and aragonite seas and the de novo acquisition of

- carbonate skeletons. *Geobiology* **8**, 256-277. (doi:10.1111/j.1472-4669.2010.00246.x).
- [47] Cusack, M., Williams, A. & Buckman, J.O. 1999 Chemico-structural evolution of linguloid brachiopod shells. *Palaeontology* **42**, 799-840.
- [48] Zhang, Z.-L., Skovsted, C.B. & Zhang, Z.-F. 2017 A hyolithid without helens preserving the oldest hyolith muscle scars; palaeobiology of *Paramicrocornus* from the Shujingtuo Formation (Cambrian Series 2) of South China. *Palaeogeography, Palaeoclimatology, Palaeoecology*. (doi:10.1016/j.palaeo.2017.07.021).
- [49] Zhuravlev, A.Y., Wood, R.A. & Penny, A.M. 2015 Ediacaran skeletal metazoan interpreted as a lophophorate. *Proc Biol Sci* **282**, 20151860. (doi:10.1098/rspb.2015.1860).
- [50] Dzik, J. 1978 Larval development of hyolithids. *Lethaia* **11**, 293-299. (doi: 10.1111/j.1502-3931.1978.tb01884.x)
- [51] Dzik, J. 1980 Ontogeny of *Bactrotheca* and related hyoliths. *Geologiska Föreningen i Stockholm Förhandlingar* **102**, 223-233. (doi:10.1080/11035898009455162).
- [52] Bengtson, S., Conway Morris, S., Cooper, B.J., Jell, P.A. & Runnegar, B.N. 1990 Early Cambrian fossils from South Australia. *Memoirs of the Association of Australasian Palaeontologists* **9**, 1-364.
- [53] Butterfield, N.J. & Nicholas, C.J. 1996 Burgess Shale-type preservation of both non-mineralizing and 'shelly' Cambrian organisms from the Mackenzie Mountains, northwestern Canada. *Journal of Paleontology* **70**, 893-899. (doi:

- 10.1017/S0022336000038579)
- [54] Freeman, G. & Lundelius, J. 2005 The transition from planktotrophy to lecithotrophy in larvae of Lower Palaeozoic Rhynchonelliform brachiopods. *Lethaia* **38**, 219-254. (doi:10.1080/00241160510013330).
- [55] Curry, G.B., Ansell, A.D., James, M. & Peck, L. 1989 Physiological constraints on living and fossil brachiopods. *Transactions of the Royal Society of Edinburgh: Earth Sciences* **80**, 255-262. (doi:10.1017/s0263593300028698).
- [56] Brown, J.H., Gillooly, J.F., Allen, A.P., Savage, V.M. & West, G.B. 2004 Toward a metabolic theory of ecology. *Ecology* **85**, 1771–1789. (doi: 10.1890/03-9000)
- [57] Bambach, R.K. 1993 Seafood through time: changes in biomass, energetics, and productivity in the marine ecosystem. *Palaeobiology* **19**, 372–397. (doi: 10.1017/S0094837300000336)
- [58] Wood, R. & Zhuravlev, A.Y. 2012 Escalation and ecological selectivity of mineralogy in the Cambrian Radiation of skeletons. *Earth Science Reviews* **115**, 249-261. (doi:10.1016/j.earscirev.2012.10.002).
- [59] Peck, L.S. 1992 Body volumes and internal space constraints in articulate brachiopods. *Lethaia* **25**, 383–390. (doi: 10.1111/j.1502-3931.1992.tb01641.x)
- Figure 1.** *Pedunculotheca diania* Sun, Zhao et Zhu gen. et sp. nov. from the Chengjiang Biota, Yunnan Province, China. Fossil images in the bottom row

correspond to their interpretive drawings in the top row. (a) NIGPAS 166593, holotype, complete specimen with operculum and attachment structure with terminal holdfast; detail of boxed region shown in (b). (c) NIGPAS 166594, dorsoventrally compacted specimen with operculum and partial pedicle structure. Detail of boxed region shown in (d). Scale bar = 2 mm. Abbreviations: cc = coelomic cavity, co = conical shell, da = dorsal apex, do = dorsum, ho = holdfast, op = operculum, sp = spine, ve = venter.

Figure 2. *Pedunculotheca diania* Sun, Zhao et Zhu gen. et sp. nov. from the Chengjiang Biota, Yunnan Province, China. (a) NIGPAS 166595, shell with incomplete pedicle structure with coelomic cavity. Detail of boxed region shown in (b). (c) NIGPAS 166599a, juvenile conical shell with operculum showing two longitudinal ventral grooves and circular larval shell. (d) NIGPAS 166598, interior of external mould of operculum displaying circular outline. (e) NIGPAS 166597, a conical shell preserved operculum and soft parts, showing a compressed elliptic cross-section. (f) NIGPAS 166600, juveniles showing intact larval shells and two or one central longitudinal grooves; detail of box shown in (g). Scale bars = 2mm for (a, c, e and f) and 1mm for (b, d and g). Abbreviations: cc = coelomic cavity, da = dorsal apex, pe = pedicle.

462

463 **Figure 3.** Artistic reconstruction of *Pedunculotheca diania* Sun, Zhao et Zhu gen. et
464 sp. nov. Adult specimen shown in feeding position.

465

466 **Figure 4.** Majority rule consensus of all trees that are optimal under parsimony
467 analysis using equal or implied weights ($3 \leq k \leq 24$), indicating key evolutionary
468 transitions. Node labels denote frequency in optimal trees (first figure) and posterior
469 probability under Bayesian analysis (second figure). For details, see Supplementary
470 information.